

93. (±)-4-Amino-4,5-dideoxyribose, (±)-4-Amino-4-deoxyerythrose, and (±)-Dihydroxyproline Derivatives from *N*-Dienyl- γ -lactams

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Hetero-*Diels-Alder* cycloaddition of acylnitroso dienophile **4** with the *N*-(butadienyl)pyrrolidinone derivatives **2a, b** led with complete regioselectivity to the oxazine adducts **5a, b** (*Scheme 1*). Sequential osmylation, protection of the ensuing glycol, and reduction of the N–O bond gave the expected hemiaminals **11a, b** which were characterized by their crystalline sulfite adducts **12a, b** (*Schemes 1* and *2*). Deprotection and saponification of the latter led to aminodeoxyerythrose and to aminodeoxyribose derivatives as an equilibrium of pyrrolidinose equivalents, *i.e.*, hemiaminals **14a, b**, imines **14'a, b**, and dimers **14''a, b**, respectively (*Scheme 3*). Hydrocyanic acid addition to **11a, b** led ultimately to the proline derivatives **16a, b** (*Scheme 2*). Compound **11b** proved to be an inhibitor of syncytium formation in AIDS-infected cells.

Introduction. – Pyrrolidinosugars, *i.e.*, 4-amino-4-deoxyaldoses, were described for the first time by *Paulsen* and coworkers in the L-xylose [1], L-lyxose [2], and D-glucose and D-galactose [3] series; more recently, in the D-arabinose, *i.e.*, nectrisine or FR 900483 [4], and in the D-ribose series [5], they were shown to be immunomodulators and glycosidase inhibitors. In their protected imino form, the D-ribose and D-threose derivatives were used for the synthesis of C-glucosides [6] and of 3,4-dihydroxyprolines [7]. Clearly these pyrrolidinoses are of interest as potential bio-active molecules and as synthetic intermediates.

Some time ago, we described the total synthesis of some racemic 4-amino-4-deoxyerythrose derivatives, the key step being hetero-*Diels-Alder* cycloadditions of acylnitroso dienophiles of type **4** with 1-(silyloxy)butadiene **1** (followed by *cis*-hydroxylation and reduction) [8]. In this case, the hetero-*Diels-Alder* cycloaddition proved to be non-regiospecific.

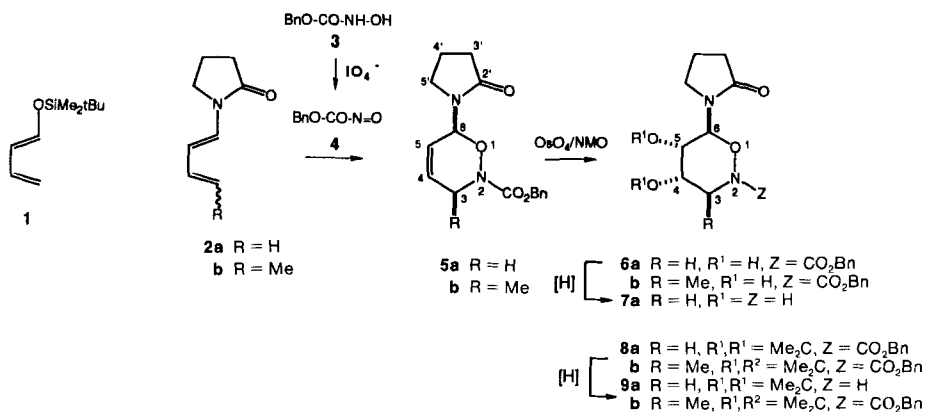
We describe herein the total synthesis of two racemic pyrrolidinose derivatives, *i.e.*, of 4-amino-4-deoxyerythrose **14a** and of 4-amino-4,5-dideoxyribose **14b**, using a similar methodology starting from the known *N*-dienylpyrrolidinones **2a** and **2b** [9], respectively. The hetero-*Diels-Alder* cycloaddition proved to be regiospecific. Acylnitroso dienophiles of type **4** are highly reactive species which cannot be isolated; they were prepared *in situ* in the presence of the diene partners by oxidation of the hydroxamic acids of type **3** with an appropriate periodate [10a, b].

We also describe the facile synthesis of the racemic dihydroxyprolines **16a** and **16b** simply by adding hydrocyanic acid to the protected pyrrolidinoses [1] [11]. Dihydroxyproline **16a** had already been synthesized, both as a racemic mixture [12] and as a chiral

entity [13], and is a potential glycosidase inhibitor: its all-*trans* (2*R*,3*R*,4*R*)-isomer was shown to be a potent inhibitor of β -D-glucuronidase [14].

Hetero-Diels-Alder Cycloaddition and Osmylation. – Dienyllactams **2a** and **2b** were prepared according to a known procedure by condensation of pyrrolidin-2-one to crotonaldehyde or to pentenal in toluene solution, H₂O being eliminated by azeotropic distillation [9]. Hetero-Diels-Alder cycloaddition was performed with these dienes at 0° in CH₂Cl₂/MeOH in the presence of hydroxamic acid **3** which was oxidized *in situ* with (BnMe₃N)IO₄ [10c] (Scheme 1). The reaction proved to be regiospecific; in both cases, only one adduct was formed, *i.e.*, **5a** and **5b**, respectively.

Scheme 1



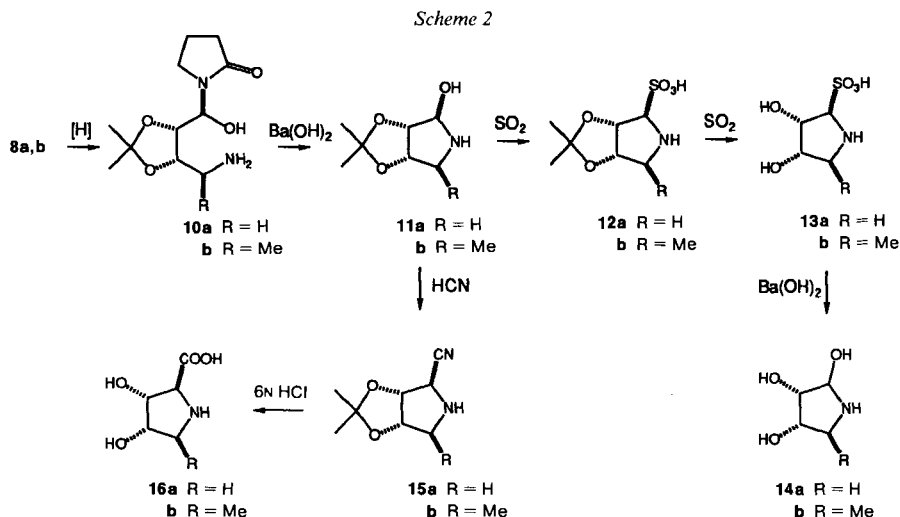
Diene **2b** is a mixture of two geometrical diastereoisomers, *i.e.*, of the (1*E*,3*E*)- and (1*E*,3*Z*)-isomers whose ratio depends on the reaction conditions: it ranged from *ca.* 1:1 before workup to 1:3 after chromatography; in the crystalline form only the (1*E*,3*Z*)-isomer was present. Whatever the ratio of the (*E,E*)/(*E,Z*)-mixture, only *cis*-cycloadduct **5b** was formed in 50–60% yield. When the reaction was monitored by ¹H-NMR, a fast (albeit incomplete) (*E,Z*)→(*E,E*) isomerization was observed during the addition of some impure tetraalkylammonium periodate (*i.e.*, slightly colored by the presence of I₂) to the solution of **2b** in CDCl₃, the (*E,E*)-isomer becoming the major product. Next, hydroxamic acid **3** was added to the reaction medium: the (*E,E*)-diene reacted at a high rate, while the (*E,Z*)-isomer disappeared slowly. Once the reaction was complete, diene **2b** had totally disappeared, and cycloadduct **5b** was the only product. We could verify by ¹H-NMR that the (*E,Z*)→(*E,E*) isomerization does not operate, neither spontaneously, nor in the presence of hydroxamic acid **3**, nor in the presence of *pure* tetraalkylammonium periodate. On the other hand, we verified that I₂ promotes a fast (*E,Z*)→(*E,E*) isomerization, the final (*E,E*)/(*E,Z*) composition being 4:1.

These results are best explained as follows: *cis*-cycloadduct **5b** was obtained *via* a classical (concerted) [4πs+2π] cycloaddition process of the (*E,E*)-isomer of **2b** with nitroso dienophile **4**. The (*E,Z*)-isomer, which is much less reactive toward the nitroso dienophile, isomerized gradually to the (*E,E*)-form *under the action of* I₂ [15], a species always present in the reaction medium, so that only *cis*-cycloadduct **5b** was formed. This

unexpected hetero-*Diels-Alder* reaction was to compare to the results obtained by *Zeza* and *Smith* for the cycloaddition of **2b** with ethyl acrylate which, according to these authors, led to a normal stereochemical outcome [9].

Bis-hydroxylation of adducts **5a** and **5b** was performed with catalytic amounts of osmium tetroxide in the presence of the co-oxidant *N*-methylmorpholine *N*-oxide (NMO) [16]. It led stereospecifically to the diols **6a** and **6b**, respectively, the osmylation occurring from the less hindered side, *i.e.*, *anti*; these results agree with those found previously [8] [17]. Diols **6a, b** were characterized as such and as their acetonides **8a, b**.

Reduction Processes to Aminodeoxysugar Derivatives (Scheme 2). – The reductive cleavage of the cyclic diols was studied in the simple series (R=H). It was found that catalytic hydrogenolysis (over Pd/C) in MeOH proceeded in two steps: *i*) debenzoylation and deprotection of the N-atom proceeded at high rate at room temperature and led to oxazine **7a**; *ii*) hydrogenolysis of the N–O bond occurred at 40° and led directly to the



pyrrolidine-3,4-diol, as already observed in the silyloxy series [8]; nevertheless, the expected intermediate aminosugars could not be trapped. Therefore, we turned our attention to the reductive cleavage of the acetonide derivative **8a** by applying the same methodology as described above. Once again, the deprotection of the N-atom proceeded at high rate, leading to the expected oxazine **9a** which, after 1 day under the same reaction conditions, led quantitatively to the hemiaminal **10a**, *i.e.*, a simple derivative of 4-amino-4-deoxyerythrose. This compound proved to be a stable entity which was characterized as its picrate salt and as its diacetyl derivative.

Hydrogenolysis of **8b** led to the oxazine **9b** whose reduction with *Raney*-Ni gave crystalline hemiaminal **10b**. Compounds **10a** and **10b** are stable species in neutral and in acidic medium. In basic medium ($Ba(OH)_2/H_2O$), and partially by thermolysis, they lead to pyrrolidinone and to the pyrrolidinoses **11a** and **11b**, respectively. These aminosugars were easily transformed into their crystalline sulfonates (sulfite adducts) **12a** and **12b** by

the action of gaseous SO_2 . Further reaction of SO_2 led to cleavage of the acetonides and to the formation of the crystalline sulfonates **13a** and **13b** which were saponified by $\text{Ba}(\text{OH})_2$ to the aminosugars **14a** and **14b**, respectively.

These 'free' aminosugars **11a, b** and **14a, b** are tautomeric mixtures in water solution (D_2O ; see *Table 1* and *Scheme 3*). The acetonide-protected aminosugars occur as a mixture of the hemiaminal **11a, b** and of the corresponding imine form **11'a, b** (formed by dehydration). The aminosugars having free OH groups occur as mixtures of three components, the hemiaminal **14a, b**, the corresponding imine form **14'a, b**, and one dimer **14''a, b** – probably 'cis-syn-cis' for **14''a**, and 'cis-anti-cis' for **14''b** – which is favored at high concentration or in the absence of any solvent. Dissolution of hemiaminals of type **11** or **14** does not lead immediately to the equilibrium, as observed in particular for **14a**. In all instances, an increase of temperature favors the imine forms **11'a, b** and **14'a, b**; this is particularly clear for **14a**. The acetonides **11a, b** are soluble in organic solvents and can be extracted from their aqueous solution by Et_2O , occurring thereby only in their imine forms **11'a, b** (as determined by $^1\text{H-NMR}$ in CDCl_3).

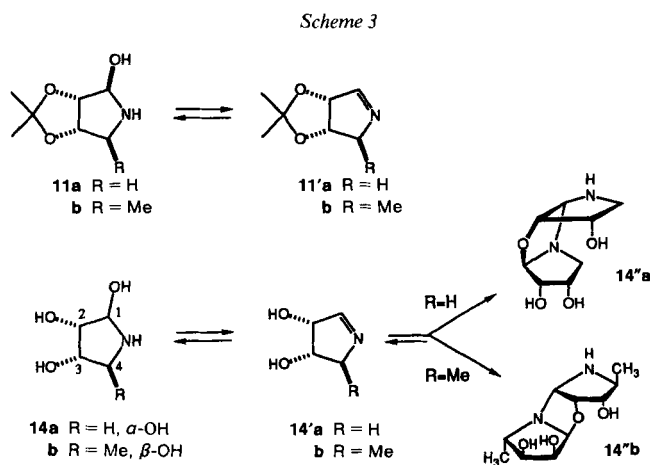


Table 1. Proportions of Tautomeric Structures (see Scheme 3) of the Aminosugars 11a, b and 14a, b in Solution in D_2O ($^1\text{H-NMR}$ determination)^{a)}

	Temperature [K]	Imine form [%]	Hemiaminal form [%]	Dimer [%]
11a	300	40	60	–
	332	55	45	–
11b	300	80	20	–
14a	300 ^{b)}	42	25	33
	300 ^{c)}	18	71	11
	320	36	48	16
	343	ca. 80	ca. 10	ca. 10
	300	12	20	68
14b	320	26	23	51
	340	46	27	27

^{a)} Concentrations: **14a**, 0.15M; **14b**, 0.26M; **11a, b**, ca. 0.2M. ^{b)} Immediately after solubilization. ^{c)} After 15 h in solution.

This type of equilibrium seems to be a general phenomenon with pyrrolidinoses and was already described qualitatively by *Paulsen* and coworkers in the L-lyxose and in the L-xylose series; these authors demonstrated in particular the structure of the corresponding dimers [1] [2]. In the D-arabinose series, a Japanese group cited the imine form only for nectrisine [4]. In the D-ribose series (4-amino-4-desoxy-D-ribose), *Witte* and *McClard* discussed the equilibrium between the various species (hemiaminal, imine, and dimer) as a function of pH [5].

Conversion to the Racemic Amino Acids 16a and 16b. – The addition of hydrocyanic acid to aminosugars in their hemiaminal form is a reaction of wide scope which was described in aqueous medium by *Böshagen et al.* for nojirimycine (5-amino-5-desoxy-D-glucose) [11] and by *Paulsen* and coworkers for L-xylopyrrolidinose [1] and for D-xyloperidinose [18]. This type of addition was also described in anhydrous medium using Me_3SiCN as cyanating agent [7].

The addition of HCN to the acetonides **11a** and **11b** in Et_2O solution led stereospecifically to the aminonitriles **15a** and **15b** (Scheme 2), respectively, the approach of the CN^- anion occurring *trans* to the acetonide ring. Acid hydrolysis of the nitrile function to the carboxylate was performed with 6N HCl during 1–2 days at 80° according to *Arakawa* and *Yoshifuji* [7]. It led in good yield to the corresponding dihydroxyprolines **16a** and **16b** as the only reaction products; **16a** is a known racemic product [12]; the spectroscopic data were identical with those reported [12] [13a, c].

Structural and Conformational Analyses. – *Cycloadducts 5a, b and Diols 6a, b.* Cycloadducts **5a, b** are 3,6-dihydro-2H-oxazines whose conformation and relative configuration could easily be deduced from the magnitude of the vicinal and allylic coupling constants between H–C(3) and H–C(6) and the olefinic H–C(4) and H–C(5) [8] [17] [19] (see Table 2). *E.g.*, for adduct **5b**, the relatively large $J(3,4)$ and the rather small $J(5,6)$ indicate that H–C(3) is strictly pseudoequatorial and H–C(6) strictly pseudoaxial, the conformation being typically half-chair. This clearly points to a *cis*-topology of the substituents at C(3) and C(6).

As to the diols **6a, b**, the large magnitude of $J(5,6)$ is in good agreement with H–C(5) and H–C(6) being *trans*-diaxial in a typical chair conformation. The small magnitude of $J(4,5)$ points to an equatorial H–C(4). From these data it follows that the two OH groups are in *cis*-topology and both *trans* with respect to the pyrrolidinone moiety. In **6b**, Me–C(3) is axial and the pyrrolidinone moiety equatorial. It is worth noticing that H_a –C(3) and H_b –C(3) of **6a** (also of **8a**) have very differentiated chemical shifts; this is due to the anisotropy of the *N*-acyl moiety which deshields the equatorial H_a –C(3) (4.5 ppm) by ca. 1 ppm with respect to the axial H_b –C(3) (3.6 ppm) [20] (see also [8] [17]). This pronounced chemical-shift difference disappears in the *N*-non-acylated compounds **7a** and **9a** which nevertheless have the same conformation as those described above.

Linear Aminosugars 10a, b. The medium values (5–9 Hz) of all the vicinal coupling constants between the protons of the aminosugar moiety indicate a linear rather than a cyclic structure. The existence of the NH_2 group, only present in a linear compound, is demonstrated for **10a** by the formation of a diacetyl derivative containing the amide group NHAc, whose well-defined NH signal in the $^1\text{H-NMR}$ spectrum is coupled with both neighboring H–C(4) (see *Exper. Part*).

Aminosugars and Cyanohydrins. These five-membered rings (pyrrolidines) are known to have rather flexible conformations. Their vicinal *cis*- and *trans*-coupling constants cannot easily be differentiated since J_{cis} varies from 4 to 7 Hz and J_{trans} from 0 to 9 Hz [21]. Therefore, the configuration at C(1) is difficult to ascertain (Table 3). Nevertheless, should an anomeric effect exist, *e.g.* in furanoses, a single rule seems to be valid, *i.e.*, $J_{trans}(1,2)$ ca. 0–2 Hz and $J_{cis}(1,2) = 4$ –5 Hz [21]. From the observed small $J(1,2)$ values, it follows that pyrrolidinoses **11a, b** and **14b** and cyanohydrins **15a, b** occur as *trans*-isomers (β -anomers for aminosugars), whereas aminoerythrose **14a** appears as the *cis*-isomer (α -anomer) ($J_{cis}(1,2) = 5.8$ Hz). This latter conclusion is confirmed by the observed shielding of H–C(1) and H–C(2) with respect to the corresponding H-atoms of **14b** whose H–C(1) (resp. H–C(2)) is deshielded by the neighboring OH–C(2) (resp. OH–C(1)) in *cis*-position.

The dimerization in the ribose series led to dimer **14''b** whose $^1\text{H-NMR}$ data pertaining to the anomeric H-atoms of both pyrrolidine moieties are similar to those reported for a dimer belonging to the L-lyxose series [1].

Table 2. $^1\text{H-NMR}$ Data (CDCl_3) of Oxazines 5–9. 250 MHz, 300 K; δ in ppm, J in Hz, int. standard SiMe_4^a .

	$\text{H}_a\text{-C}(3)$	$\text{H}_b\text{-C}(3)^b$	H–C(4)	H–C(5)	H–C(6)	PhCH_2^c	$J(3a,3b)^b$	$J(3a,4)$	$J(3b,4)$	$J(4,5)$	$J(5,6)$
5a ^{d)}	4.14	4.14	6.13	5.70	6.13	5.16, 5.28	n.d.	3.2	–	10.2	2.5
b ^{f)}	4.55	1.35	6.07	5.57	6.27	5.18, 5.28	6.7	4.6	–	10.4	1.6
6a	4.30	3.40	4.15	3.95	5.49	5.15, 5.23	14.4	2.6	1.4	3.0	10.0
b ^{e)}	4.52	1.31	4.01	4.37	5.18	5.18, 5.22	7.2	2.0	–	3.2	9.6
7a	3.32	3.16	4.12	3.75	5.34	–	14.5	1.6	2.7	3.2	9.6
8a ^{d)}	4.42	3.68	4.38	4.52	5.34	5.19, 5.26	14.8	2.1	3.2	5.1	8.4
b ^{h)}	4.70	1.37	4.16	4.60	5.34	5.18, 5.29	7.2	1.3	–	4.9	8.8
9a ^{h)}	ca. 3.5	ca. 3.5	4.35	4.25	5.24	–	n.d.	2.4	–	5.0	8.7
b ^{h)}	3.27	1.21	4.08	4.24	5.25	–	7.1	3.6	–	5.1	6.6

a) Pyrrolidinone moiety: 2.30–2.47 (*m*, 2 H–C(3¹)); 1.87–2.07 (*m*, 2 H–C(4¹)); 3.37–3.63 (*m*, 2 H–C(5¹)); arom. protons: 7.3–7.5.

b) For **5b**, **6b**, and **9b**, Me–C(3) instead of H_b–C(3), and $J(3a, \text{Me})$ instead of $J(3a, 3b)$.

c) $J(\text{CH}_2) = 12.3$.

d) J values in C_6D_6 .

e) $J(3,5) = 2.2$, $J(3,6) = 2.5$, $J(4,6) = 1.8$.

f) $J(3,5) = 1.6$, $J(3,6) = 2.4$, $J(4,6) = 2.0$.

g) 330 K; $\delta(\text{OH-C}(4))$: 2.61, $\delta(\text{OH-C}(5))$: 3.81; $J(4, \text{OH-C}(4)) = 1.8$, $J(5, \text{OH-C}(5)) = 6.2$.

h) Me signals of the acetamide moiety: **8a**: 1.35, 1.40; **8b**: 1.33, 1.38; **9a**: 1.40, 1.59; **9b**: 1.31, 1.50.

i) NH at 5.44.

Table 3. ¹H-NMR Data of Aminoglycoses **14a, b** and Derivatives **11a, b, 12a, b, 13a, b, and 15a, b** in D₂O. δ in ppm, J in Hz, int. ref. (D₄)-TSP or δ(DOH) = 4.76 ppm.

	H–C(1)	H–C(2)	H–C(3)	H _a –C(4)	H _b –C(4) ^{a)}	J(1,2)	J(2,3)	J(3,4a)	J(3,4b)	J(4a,4b) ^{a)}
11a ^{d)}	4.82	4.57	4.89	3.14	3.03	0	5.6	4.2	0	12.6
11'a ^{b)} ^{d)}	7.68	5.29	4.93	4.01	4.01	0	5.7	2.6	2.6	–
11b ^{d)}	4.90	4.62	4.53	3.33	1.20	1.8	6.2	2.0	–	7.0
11'b ^{c)} ^{d)}	7.62	5.39	4.58	4.27	1.20	0	5.5	0	–	7.3
12a ^{d)}	4.62	5.23	5.15	3.70	3.55	1.3	5.6	4.4	0.5	13.1
12b ^{d)}	4.54	5.17	4.74	3.88	1.44	4.3	5.8	3.1	–	7.2
13a	4.41	4.60	4.47	3.55	3.45	7.1	4.1	3.6	2.2	12.6
13b	4.39	4.58	4.04	3.67	1.42	4.6	4.8	6.7	–	6.8
14a	3.13	3.92	4.28	3.44	2.45	5.8	6.8	7.0	4.6	10.3
14'a ^{b)}	7.65	4.72	4.28	3.84	3.84	0.8	5.4	3.3	3.3	–
14'a	4.70	4.45	4.10	3.08	2.58	4.8	5.0	6.7	9.2	10.6
	4.88	4.18	4.33	3.17	2.99	1.6	4.8	4.2	5.5	11.3
14b	4.06	4.35	3.71	3.20	1.21	2.6	5.4	8.2	–	6.4
14'b ^{c)}	7.63	4.83	3.95	4.06	1.20	0.6	5.4	1.8	–	7.0
14'b	4.72	4.62	3.53	2.94	1.18	4.9	4.9	9.4	–	6.2
	5.05	4.18	3.70	2.94	1.22	4.1	5.2	8.2	–	6.2
15a ^{d)} ^{e)}	4.06	4.85	4.81	3.25	3.00	0	5.4	0	3.3	13.4
15b ^{d)} ^{e)} ^{f)}	3.99	4.97	4.50	3.47	1.27	1.8	5.6	1.3	–	7.3

a) For the 4-methyl derivatives Me–C(4) instead of H_b–C(4) and J(4a, Me–C(4)) instead of J(4a, 4b).

b) J(1,4a) = J(1,4b) = 2.5, J(2,4a) = J(2,4b) = 1.0.

c) J(1,4a) = 1.7, J(2,4a) = 1.1.

d) Acetonide signals: **11a**: 1.36, 1.48; **11'a**: 1.40, 1.41; **11b**: 1.36, 1.51; **11'b**: 1.41; **12a**: 1.39, 1.54; **12b**: 1.56, 1.40; **15a**: 1.45, 1.32; **15b**: 1.47, 1.31.

e) For convenience, **15a, b** are numbered like **11–14**; for systematic names, see *Exper. Part*.

f) In CDCl₃, ref. SiMe₄; δ(NH) = 2.56; J(1, NH) = 5.8, J(4, NH) = 4.0.

It follows that **14'b** is likely to occur in the 'cis-anti-cis'-topology postulated for this lyxose dimer. On the contrary, for the dimer **14'a**, one J(1,2) is small; a simple explanation is that this dimer appears in the 'cis-syn-cis'-topology (Scheme 3).

In the sulfite adducts **12a, b**, and **13a, b**, the anomeric effect (of SO₃H) is known to be weak or even inverse, and the SO₃H group appears in its equatorial configuration [22] [23]. As a consequence, the J(1,2) values vary widely. The trans-configuration was demonstrated for **13b** by nuclear Overhauser effects (NOE, H–C(1) and H–C(4) being cis to each other: irradiation of Me–C(4) led to a NOE of 6% for H–C(2) and of 11% for H–C(3); and irradiation of H–C(4) to a NOE of 6% for H–C(1).) The configuration of the other sulfite adducts is β-DL also since J(1,2) is small for **12a** and since **12b** and **13a** are directly related to the preceding ones.

AIDS-Inhibition Assays. – Some of the compounds synthesized herein were submitted to anti-HIV assays (see *Exper. Part* and Table 4). The aminodeoxyribose **11b** proved

Table 4. Anti-HIV Activities and Cytotoxicities as Determined for Compounds **7a, 11b, and 13a, b** (in μM)

	Syncytia inhibition	EC ₅₀ ^{a)}	TC ₅₀ ^{b)}
7a	> 10000	> 10000	> 10000
11b	> 100	20	100
13a	> 2000	400	4000
13b	> 2000	400	4000
AZT	> 0.4	0.016	> 1000

a) Concentration which reduces the virus yield by 50% in infected cells.

b) Concentration which reduces cell growth by 50%.

to be somewhat more active than castanospermine [24], but less active than AZT. Nevertheless, **11b** showed a relatively pronounced cytotoxicity.

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Experimental Part

1. *General. Raney-Ni* (slurry in H₂O), 5% Pd/C catalyst, 2,2-dimethoxypropane, pent-2-enal, *N*-methylmorpholine-*N*-oxide (NMO), OsO₄, *tert*-butyl hydroperoxide, SO₂ gas, pyrrolidin-2-one, and toluene-4-sulfonic acid (TsOH) were purchased from *Fluka*, crotonaldehyde from *Merck*, and NaCN and Ba(OH)₂·8 H₂O from *Prolabo*. Usual solvents were freshly distilled, CH₂Cl₂ was kept over Na₂CO₃. Flash chromatography (FC): silica gel (*Merck* 60, 230–400 mesh). TLC: Al-roll silica gel (*Merck* 60, F₂₅₄). M.p.: *Kofler* hot-bench or *Büchi-SMP-20* apparatus; corrected. IR Spectra (ν in cm⁻¹): *Perkin-Elmer 157 G* or *Nicolet 205*. ¹H- and ¹³C-NMR Spectra: *Bruker AC-F250*, using double-irradiation techniques; SiMe₄ or sodium (trimethylsilyl)(D₄)propanoate ((D₄)-TSP) in D₂O (¹H-NMR), and CDCl₃, C₆D₆, CD₃OD, or (in D₂O) CH₃OH or dioxane (δ (CDCl₃) 77.0, δ (C₆D₆) 128.0, δ (CD₃OD) 49.0, in D₂O δ (CH₃OH) 50.3, δ (dioxane) 67.6 with respect to SiMe₄; ¹³C-NMR) as internal references; δ in ppm and *J* in Hz. High resolution (HR) MS were measured on a *MAT-311* spectrometer at the University of Rennes. Microanalyses were carried out by the Service Central de Microanalyses of the *CNRS*, F-69390 Vernaison.

2. *Dienylpyrrolidinones. N-(Buta-1,3-dienyl)pyrrolidin-2-one (2a)*. According to [9] with some modifications: A soln. of pyrrolidin-2-one (20 g, 18 ml, 0.234 mol), TsOH (20–100 mg), and crotonaldehyde (19.6 ml, 0.24 mol, 1 equiv.) in toluene (100 ml) was refluxed and the H₂O removed in a *Dean-Stark* trap. After 1 h and again after 1.5 h, crotonaldehyde (9.8 ml (0.5 equiv.) and 4.9 ml (0.25 equiv.)) was added. After 2 h, the cooled soln. was washed with aq. 1M Na₂CO₃, H₂O, and brine (3 × 20 ml). The aq. phases were extracted with Et₂O (2 × 50 ml) and the org. phases dried (MgSO₄) and evaporated. FC (Et₂O) gave **2a** (18.0 g, 56%) as a yellow resin which crystallized at 0°. Yellow crystals. M.p. 58° ((*i*-Pr)₂O) ([9]: 58–59°). IR (KBr): 1695, 1645, 1430, 1395, 1300. ¹H-NMR (CDCl₃): 2.13 (*q*, *J* = 8, 2 H–C(4)); 2.51 (*t*, *J* = 8, 2 H–C(3)); 3.56 (*t*, *J* = 7, 2 H–C(5)); 4.99 (*d*, H_a–C(4'')); 5.14 (*d*, H_b–C(4'')); 5.63 (*dd*, H–C(2'')); 6.35 (*dt*, H–C(3'')); 7.11 (*d*, H–C(1'')); *J*(1',2') = 14.3, *J*(1',3') = *J*(1',4'a) = *J*(1',4'b) = 0.8, *J*(2',3') = 10.4, *J*(2',4'a) = *J*(2',4'b) = 0.8, *J*(3',4'a) = 10.3, *J*(3',4'b) = 16.9, *J*(4'a,4'b) = 1.7.

N-(Penta-1,3-dienyl)pyrrolidin-2-one (2b). A soln. of pyrrolidin-2-one (8.0 ml, 0.1 mol), TsOH (23 mg) and pent-2-enal [25] (5.2 ml, 53 mmol) in toluene (67 ml) was refluxed for 3 h as above. The cooled soln. was washed with aq. 1M Na₂CO₃ and H₂O (2 × 20 ml) and the aq. soln. extracted with Et₂O (2 × 50 ml). The org. solns. were dried (MgSO₄) and evaporated: **2b** (7.2 g, 90%) (*E,E*)/(*E,Z*) 1:1. Purification by FC (Et₂O) gave **2b** (4.97 g, 62%), (*E,E*)/(*E,Z*) 1:3 ([9]: 81:19 mixture), crystallizing at 0° as the (*E,Z*)-isomer. Yellow resin (isomer mixture) or yellow crystals. M.p. 70° (toluene/cyclohexane). ¹H-NMR (CDCl₃): 2.11 (*m*, 2 H–C(4)); 2.51 (*m*, 2 H–C(3)); 3.60 (*m*, 2 H–C(5)); (*E,Z*)-isomer: 7.07 (H–C(1')); 5.81 (H–C(2')); 6.02 (H–C(3')); 5.43 (H–C(4')); 1.73 (Me–C(4')); *J*(1',2') = 14.0, *J*(1',4') = 0.9, *J*(2',3') = 11.0, *J*(2',4') = 0.9, *J*(3',4') = 10.8, *J*(3',Me) = 1.7, *J*(4',Me) = 7.2; (*E,E*)-isomer: 6.96 (H–C(1')); 5.58 (H–C(2')); 6.03 (H–C(3')); 5.60 (H–C(4')); 1.74 (Me–C(4')); *J*(1',2') = 14.4, *J*(1',4') = 0.6, *J*(2',3') = 10.4, *J*(2',4') = 0.6, *J*(3',4') = 15.1, *J*(3',Me) = 1.7, *J*(4',Me) = 6.6. ¹³C-NMR: identical to [9].

3. *Diels-Alder Addition. General Procedure*. To a soln. of diene **2** (10 mmol) in 10 ml of solvent containing (BnMe₃N)IO₄ [10c] (1.37 g, 4 mmol) in an ice bath was added within 0.5 h portionwise hydroxamic acid **3** [8] (2.0 g, 12 mmol, 1.2 equiv.). After 1 h at 0°, AcOEt (50 ml) was added to the red soln. and the org. phase washed with 1M aq. Na₂CO₃ (10 ml; and enough Na₂SO₃ to suppress color) and brine (3 × 10 ml). The aq. phases were extracted with AcOEt and the combined org. soln. dried (MgSO₄) and evaporated to give crude adducts **5**.

(±)-*Benzyl 3,6-Dihydro-6-(2'-oxopyrrolidin-1'-yl)-2H-1,2-oxazine-2-carboxylate (5a)*. *General Procedure* with **2a** (13.55 g, 99 mmol), (BnMe₃N)IO₄ (13.55 g, 39.5 mmol) in CH₂Cl₂ (48 ml)/MeOH (48 ml), and **3** (19.8 g, 119 mmol, 1.2 equiv.). Crude product was washed with Et₂O and recrystallized: **5a** (12.5 g, 42%). FC of the mother liquor gave a second crop (1.5 g, 5%). Cream-colored crystals. M.p. 90° (AcOEt/Et₂O). IR (KBr): 2965, 1785, 1455, 1435, 1405, 1360, 1280, 1270, 1220, 1210, 1090, 1010, 700. ¹H-NMR: *Table 2*. ¹³C-NMR (CDCl₃): 175.5 (C(2)); 154.9 (NCO₂); 135.9, 128.4, 128.1, 128.0 (arom. C); 127.5, 123.5 (C(4), C(5)); 77.7 (C(6)); 67.5 (PhCH₂); 44.2 (C(3)); 44.1 (C(5')); 30.8 (C(3')); 18.2 (C(4')). Anal. calc. for C₁₆H₁₈N₂O₄ (302.33): C 63.56, H 6.00, N 9.27; found: C 63.4, H 5.9, N 9.3.

(±)-Benzyl 3,6-Dihydro-r-3-methyl-c-6-(2'-oxopyrrolidin-1'-yl)-2H-1,2-oxazine-2-carboxylate (**5b**). *General Procedure* with **2b** (4.44 g, 29.2 mmol), (BnMe₃N)IO₄ (4.03 g, 10.7 mmol), and **3** (5.89 g, 35.2 mmol, 1.2 equiv.) in MeOH (15 ml)/CH₂Cl₂ (15 ml). The crude product (9.84 g) was purified by FC (AcOEt, 180 g of SiO₂): pure **5b** (5.82 g, 63%). Yellowish resin. ¹H-NMR: *Table 2*. ¹³C-NMR (CDCl₃): 175.7 (C(2')); 154.7 (NCO₂); 135.9, 128.4, 128.0, 127.9 (arom. C); 133.4 (C(4)); 123.7 (C(5)); 77.9 (C(6)); 67.6 (PhCH₂); 49.8 (C(3)); 42.4 (C(5')); 31.0 (C(3')); 17.9, 17.7 (C(4'), Me-C(3)). Too unstable for analysis.

4. *Cyclic Diols. General Procedure for cis-Hydroxylation* [8] [16]. The catalyst was prepared according to [26] from OsO₄ (1 g) and 1 ml of 70% *t*-BuOOH in *t*-BuOH (200 ml). To a stirred soln. of **5** (10 mmol) and *N*-methylmorpholine *N*-oxide (NMO; 2.0 g, 15 mmol, 1.5 equiv.) in acetone (10 ml) and H₂O (5 ml) was added dropwise a soln. of OsO₄ (3 ml). The soln. was kept 17–24 h at 40° until completion of the reaction. AcOEt (50 ml) was added, the org. phase washed with brine (3 × 10 ml), the aq. phase extracted with AcOEt, and the combined org. soln. dried (MgSO₄) and evaporated.

(±)-Benzyl *r*-4,*c*-5-Dihydroxy-*t*-6-(2'-oxopyrrolidin-1'-yl)-1,2-oxazine-2-carboxylate (**6a**). *General Procedure* with **5a** (12.45 g, 41.2 mmol), NMO (8.36 g, 62 mmol), acetone (38 ml), H₂O (21 ml), and OsO₄ soln. (12.5 ml; 17 h at 40°). The diol crystallized and was washed with *i*-PrOH: pure **6a** (12.22 g, 88%). Colorless crystals. M.p. 154° (EtOH). IR (KBr): 3440, 3310, 2930, 1705, 1675, 1412, 1345, 1335, 1285, 1275, 1215, 1090, 1000, 985, 752, 692. ¹H-NMR: *Table 2*. Anal. calc. for C₁₆H₂₀N₂O₆ (336.34): C 57.13, H 5.99, N 8.33; found: C 57.2, H 6.1, N 8.3.

(±)-Benzyl *t*-4,*t*-5-Dihydroxy-*r*-3-methyl-*c*-6-(2'-oxopyrrolidin-1'-yl)-1,2-oxazine-2-carboxylate (**6b**). *General Procedure* with **5b** (21.3 g, 67.4 mmol), NMO (18.19 g, 135 mmol), acetone (60 ml), H₂O (30 ml) and OsO₄ soln. (25 ml): **6b** (19.25 g, 81%). Oil. Characterized as its acetone **8b**. ¹H-NMR: *Table 2*.

(±)-*r*-4,*c*-5-Dihydroxy-*t*-6-(2'-oxopyrrolidin-1'-yl)-1,2-oxazine (**7a**). A stirred soln. of **6a** (0.64 g, 1.9 mmol) in MeOH (6.4 ml) was hydrogenated over 5% Pd/C (77 mg) for 3 h at r.t. After centrifugation, the solvent was evaporated. The resulting crystals were washed with *i*-PrOH: pure **7a** (0.377 g, 98%). Colorless crystals. M.p. 190° (MeOH). ¹H-NMR: *Table 2*. ¹³C-NMR (D₂O, ref. MeOH): 182.4 (C(2')); 81.8 (C(6)); 68.0, 67.9 (C(4), C(5)); 54.2 (C(3)); 45.0 (C(5')); 32.8 (C(3')); 19.3 (C(4')). Anal. calc. for C₈H₁₄N₂O₄ (202.21): C 47.52, H 6.98, N 13.86; found: C 47.2, H 7.3, N 13.8.

5. *Acetonides. General Procedure*. To a stirred soln. of diol **6** (10 mmol) in 2,2-dimethoxypropane (10 ml) and acetone (50 ml) was added Amberlyst-15 (H⁺ form; 200 mg). After completion of the reaction, the soln. was filtered and evaporated. The product crystallized and was washed with *i*-PrOH.

(±)-Benzyl *r*-4,*c*-5-(Isopropylidenedioxy)-*t*-6-(2'-oxopyrrolidin-1'-yl)-1,2-oxazine-2-carboxylate (**8a**). *General Procedure* with **6a** (11.15 g, 33.0 mmol), dimethoxypropane (36 ml), acetone (170 ml), and Amberlyst-15 (0.7 g; 1.5 h at 35°): pure **8a** (12.3 g, 98%). Colorless crystals. M.p. 143° (*i*-PrOH). IR (KBr): 2980, 1728, 1698, 1418, 1285, 1237, 1218, 1160, 1105, 1070, 970. ¹H-NMR: *Table 2*. ¹³C-NMR (CDCl₃): 176.1 (C(2')); 155.6 (NCO₂); 135.8, 128.4, 128.4, 128.2 (arom. C); 110.5 (Me₂C); 82.2 (C(6)); 71.3, 70.4 (C(4), C(5)); 68.0 (PhCH₂); 46.2 (C(3)); 44.0 (C(5')); 31.1 (C(3')); 27.7, 26.2 (2 Me); 18.1 (C(4')). Anal. calc. for C₁₉H₂₄N₂O₆ (376.40): C 60.62, H 6.43, N 7.44; found: C 60.4, H 6.5, N 7.4.

(±)-Benzyl *t*-4,*t*-5-(Isopropylidenedioxy)-*r*-3-methyl-*c*-6-(2'-oxopyrrolidin-1'-yl)-1,2-oxazine-2-carboxylate (**8b**). *General Procedure* with **6b** (2.80 g, 8 mmol), dimethoxypropane (8.4 ml), acetone (42 ml), and Amberlyst-15 (1.168 g; 3.5 h at 35°): pure **8a** (2.15 g, 69%). Colorless crystals. M.p. 131.5° (*i*-PrOH). IR (KBr): 3400, 2985, 1710, 1695, 1428, 1395, 1323, 1287, 1220, 1110, 1088, 1068, 1030, 995, 732. ¹H-NMR: *Table 2*. ¹³C-NMR (CDCl₃): 176.1 (C(2')); 155.4 (NCO₂); 135.9, 128.4, 128.4, 128.1 (arom. C); 110.3 (Me₂C); 82.1 (C(6)); 76.1 (C(4)); 69.0 (C(5)); 67.9 (PhCH₂); 51.2 (C(3)); 43.8 (C(5')); 31.2 (C(3')); 26.3, 27.8 (Me₂C); 18.1 (C(4')); 16.0 (Me-C(3)). Anal. calc. for C₂₀H₂₆N₂O₆ (390.42): C 61.32, H 6.71, N 7.18; found: C 60.9, H 6.7, N 7.3.

(±)-*r*-4,*c*-5-(Isopropylidenedioxy)-*t*-6-(2'-oxopyrrolidin-1'-yl)-1,2-oxazine (**9a**). A stirred suspension of **8a** (3.00 g, 7.97 mmol) in EtOH (30 ml) and MeOH (15 ml) was hydrogenated over 5% Pd/C (177 mg) for 4 h at r.t. (TLC (AcOEt) monitoring). Centrifugation and evaporation gave crystals which were washed with (*i*-Pr)₂O: **9a** (1.715 g, 89%). Colorless crystals. M.p. 145° ((*i*-Pr)₂O/AcOEt 1:1). IR (KBr): 3275, 2980, 2900, 1685, 1675, 1422, 1370, 1270, 1248, 1220, 1075, 1062, 1020, 1002, 890, 850. ¹H-NMR: *Table 2*. ¹³C-NMR (CDCl₃): 176.4 (C(2')); 110.1 (Me₂C); 82.4 (C(6)); 71.5, 70.6 (C(4), C(5)); 49.2 (C(3)); 42.9 (C(5')); 31.0 (C(3')); 27.9, 26.4 (Me₂C); 18.1 (C(4')). Anal. calc. for C₁₁H₁₈N₂O₄ (242.27): C 54.53, H 7.49, N 11.56; found: C 54.7, H 7.7, N 11.3.

(±)-*t*-4,*t*-5-(Isopropylidenedioxy)-*r*-3-methyl-*c*-6-(2'-oxopyrrolidin-1'-yl)oxazine (**9b**). A stirred suspension of **8b** (1.784 g, 4.57 mmol) in MeOH (27 ml) was hydrogenated over 5% Pd/C (109 mg) for 1.5 h at r.t. The standard workup gave **9b** (1.152 g, 98%). Colorless crystals. M.p. 149° (AcOEt/*i*-Pr)₂O 1:1). IR (KBr): 3240, 2985, 2975, 2930, 2880, 1689, 1415, 1380, 1285, 1270, 1215, 1060, 990, 840. ¹H-NMR: *Table 2*. ¹³C-NMR (CDCl₃): 176.3 (C(2')); 109.6 (Me₂C); 82.1 (C(6)); 76.2 (C(4)); 70.4 (C(5)); 53.9 (C(3)); 44.4 (C(5')); 31.2 (C(3')); 28.0, 26.5 (Me₂C);

18.3 (C(4')); 16.6 (Me–C(3)). Anal. calc. for C₁₂H₂₀N₂O₄ (256.30): C 56.23, H 7.87, N 10.93; found: C 55.4, H 7.9, N 10.8.

4-Amino-1,4-dideoxy-2,3-O-isopropylidene-1-(2'-oxopyrrolidin-1'-yl)-DL-ribose (10a). a) From **9a**: A stirred suspension of **9a** (0.27 g, 1.11 mmol) in MeOH (3 ml) was hydrogenated over 5% Pd/C (34 mg) overnight at r.t. After centrifugation, evaporation at r.t. gave pure **10a** (0.27 g, quant.).

b) From **8a**: A stirred suspension of **8a** (0.82 g, 2.24 mmol) in MeOH (8 ml) was hydrogenated over 5% Pd/C (49 mg and another 55 mg after 7 h) for 24 h at r.t.: **10a** (0.56 g, quant.). Colorless oil. ¹H-NMR (C₆D₆): 5.94 (d, H–C(1)); 4.12 (dd, H–C(2)); 3.90 (m, H–C(3)); 3.46, 3.27 (2m, 2 H–C(5')); 2.80 (m, H_a–C(4)); 2.66 (m, H_b–C(4)); 2.08 (t, 2 H–C(3')); 1.38 (m, 2 H–C(4')); 1.56, 1.47 (2s, Me₂C); J(1,2) = 9.4, J(2,3) = 5.7, J(3,4a) = 8.4, J(3,4b) = 4.8, J(4a,4b) = 12.7. ¹³C-NMR (CDCl₃): 176.0, 108.7, 77.7, 75.9, 72.2, 41.8, 40.7, 31.6, 27.7, 25.5, 18.0.

Picrate of 10a: A soln. of **10a** (60 mg, 0.25 mmol) in CH₂Cl₂ (0.2 ml) was added to a soln. of picric acid (54 mg, 0.24 mmol) in EtOH (0.1 ml). Some crystals appeared by scratching, Et₂O (0.5 ml) was added and the picrate isolated as yellow crystals. M.p. 134–139°. Anal. calc. for C₁₁H₂₀N₂O₄·C₆H₃N₃O₇ (473.29): C 43.13, H 4.88, N 14.80; found: C 42.8, H 4.8, N 14.4.

Diacetate of 10a: A mixture of **10a** (60 mg, 0.25 mmol), Ac₂O (50 μl), and pyridine (0.2 ml) was left to react for 2 h. MeOH (0.2 ml) was added, and the solvents were evaporated. The diacetate was crystallized and washed with (i-Pr)₂O to give colorless crystals (34 mg, 42%). M.p. 142° (toluene). IR (KBr): 3420, 3280, 3090, 1750, 1680, 1650, 1590, 1442, 1372, 1210, 1070, 1020. ¹H-NMR (C₆D₆): 6.72 (d, H–C(1)); 5.23 (br. s. NH); 4.20 (m, H–C(3)); 4.01 (dd, H–C(2)); 3.85 (m, H_a–C(4)); 3.14, 2.93 (2m, 2 H–C(5')); 2.96 (m, H_b–C(4)); 1.95 (t, 2 H–C(3')); 1.29 (m, 2 H–C(4')); 1.69, 1.50 (2s, 2 AcO); 1.52, 1.16 (2s, 2 Me₂C); J(1,2) = 9.2; J(2,3) = 5.6; J(3,4a) = 3.2; J(3,4b) = 8.7; J(4a,4b) = 13.7; J(NH,4a) = 7.3; J(NH,4b) = 4.6. Anal. calc. for C₁₅H₂₄N₂O₆ (328.36): C 54.86, H 7.37, N 8.53; found: C 55.0, H 7.5, N 8.4.

4-Amino-1,4,5-trideoxy-2,3-O-isopropylidene-1-(2'-oxopyrrolidin-1'-yl)-DL-ribose (10b). A soln. of **9b** (1.01 g, 4 mmol) in MeOH (18 ml) was hydrogenated over Raney-Ni (2.0 g wet) overnight at r.t. After centrifugation, evaporation gave **10a** (0.98 g, 96%). Colorless crystals. M.p. 135° (AcOEt). IR (KBr): 3330, 2905, 1669, 1610, 1436, 1370, 1290, 1265, 1225, 1090, 1072, 980, 853. ¹H-NMR (CDCl₃): 5.60 (d, H–C(1)); 4.17 (dd, H–C(2)); 3.86 (m, H–C(3)); 3.61, 3.44 (2m, 2 H–C(5')); 3.27 (m, H–C(4)); 2.42 (t, J = 7, 2 H–C(3')); 2.04 (m, 2 H–C(4')); 1.38, 1.32 (2s, Me₂C); 1.31 (d, Me–C(4)); J(1,2) = 9.4, J(2,3) = 5.4, J(3,4) = 9.3, J(4,Me–C(4)) = 6.3. ¹³C-NMR (CDCl₃): 175.7 (C(2')); 108.7 (Me₂C); 81.8 (C(6)); 76.6 (C(4)); 72.5 (C(5)); 46.1 (C(3)); 41.7 (C(5')); 31.6 (C(3')); 27.7, 25.7 (Me₂C); 23.2 (C(4')); 18.1 (Me–C(4)). Anal. calc. for C₁₂H₂₂N₂O₄ (258.31): C 55.79, H 8.58, N 10.85; found: C 55.7, H 8.7, N 10.8.

6. 4-Amino-4-deoxyerythrose, 4-Amino-4,5-dideoxyribose, and Their Derivatives. 4-Amino-4-deoxy-2,3-O-isopropylidene-DL-erythrofurranose (11a). To a stirred soln. of **10a** (0.56 g, 2.18 mmol; from 0.82 g of **8a**) in H₂O (5 ml) was added Ba(OH)₂·8 H₂O (0.69 g, 2.18 mmol). After 15 min at r.t., aq. 1N H₂SO₄ (4.3 g) was added, BaSO₄ eliminated by centrifugation, the aq. soln. extracted with Et₂O (5 × 20 ml), and the org. phase washed with brine and carefully evaporated: **11a** (0.13 g, 42%). Colorless resin: mixture **11a/11'a**. ¹H-NMR: Table 3. MS: 423 (37, M₃⁺), 267 (16), 253 (15), 142 (100, MH⁺), 126 (78), 84 (57), 59 (18). HR-MS: 423.2361 (C₂₁H₃₃N₃O₆⁺, trimer of **11'a**; calc. 423.2369).

4-Amino-4,5-dideoxy-2,3-O-isopropylidene-DL-ribofurranose (11b). To a stirred soln. of **10b** (2.0 g, 7.8 mmol) in H₂O (25 ml) was added Ba(OH)₂·8 H₂O (2.46 g, 7.8 mmol, 1 equiv.). After 30 min, the centrifuged aq. soln. was extracted with Et₂O (5 × 20 ml) and the org. phase washed with brine and carefully evaporated: **11b** (0.90 g, 75%). Colorless resin: mixture **11b/11'b**. ¹H-NMR: Table 3. ¹H-NMR (CDCl₃) of **11'b**: 7.51 (d, J = 1.6, H–C(1)); 5.11 (d, J = 5.7, H–C(2)); 4.31 (m, H–C(3), H–C(4)); 1.38, 1.35 (2s, Me₂C); 1.21 (d, J = 7.4, Me–C(4)). ¹³C-NMR (CDCl₃) of **11'b**: 164.1 (C(1)); 111.7 (Me₂C); 86.3, 86.3 (C(2), C(3)); 73.6 (C(4)); 27.0, 25.7 (Me₂C); 19.3 (Me–C(4)).

4-Amino-4-deoxy-DL-erythrofurranose (14a). To a stirred soln. of **13a** (0.25 g, 1.35 mmol) in H₂O (5 ml) was added Ba(OH)₂·8 H₂O (0.43 g, 1.35 mmol, 1 equiv.). After 20 min, precipitated BaSO₃ was separated and washed with MeOH (baryte can be exactly precipitated with 1N H₂SO₄), the solns. were evaporated to give **14a**, i.e., **14a/14'a/14''a** (0.12 g, 88%). Yellowish resin. ¹H-NMR: Table 3. ¹³C-NMR (D₂O) of **14a**: 87.5, 74.0, 69.4, 55.2; other peaks: 100.5, 87.7, 81.4, 74.6, 71.8, 68.3, 60.2, 51.6, 51.4.

4-Amino-4,5-dideoxy-DL-ribofurranose (14b). By the same procedure, **14b** was prepared from **13b** or directly from **10b**. To a soln. of **10b** (0.274 g, 1.06 mmol) in H₂O (5 ml) was added Ba(OH)₂·8 H₂O (0.336 g, 1.06 mmol, 1 equiv.) and stirred for 20 min at r.t. Baryte in excess was eliminated and the soln. stirred under SO₂ atmosphere in a glass vessel overnight at 35°, then evaporated. The white solid (BaSO₃ + **13b**) was washed with MeOH (to eliminate the pyrrolidinone) and dissolved in H₂O (5 ml), and baryte (0.336 g) was added. The suspension was

stirred for 20 min and filtered and the soln. evaporated (excess of baryte can be exactly precipitated with 1N H₂SO₄) to give **14b**, i.e., **14b/14'b/14''b** (96 mg, 79%). Yellowish resin. ¹H-NMR: Table 3. ¹³C-NMR (D₂O): **14b** and **14''b**: 98.3, 81.3, 81.2, 80.0, 78.9, 77.6, 77.0, 73.7, 71.3, 63.0, 57.2, 56.9, 18.6, 18.1, 17.1; **14'b**: 169.2, 77.7, 75.3, 74.2, 18.0.

7. *Sulfite Adducts. 4-Amino-1,4-dideoxy-2,3-O-isopropylidene-DL-erythrofurano-1-sulfonic Acid (12a)*. Direct action of SO₂: A soln. of **10a** (0.578 g, 2.37 mmol) in Et₂O (5 ml) was stirred under SO₂ in a glass vessel at –20°. Some white crystals appeared immediately. After 1 h, MeOH (1 ml) was added and **12a** (0.265 g, 50%) isolated by filtration. White crystals. M.p. ca. 180° (subl.; EtOH). IR (KBr): 3430, 3080, 1645, 1615, 1448, 1388, 1377, 1275, 1255, 1225, 1165, 1070, 1055, 1035, 1010. ¹H-NMR: Table 3. ¹³C-NMR (CD₃OD): 113.7 (Me₂C); 82.2 (C(2)); 80.0 (C(3)); 77.9 (C(1)); 52.6 (C(4)); 29.1, 24.0 (Me₂C). Anal. calc. for C₇H₁₃NO₃S (223.24): C 37.66, H 5.88, N 6.27, S 14.36; found: C 37.9, H 5.8, N 6.5, S 14.0.

4-Amino-1,4,5-trideoxy-2,3-O-isopropylidene-DL-ribofuranose-1-sulfonic Acid (12b). An Et₂O soln. of **11b** was prepared from **10b** (0.40 g, 1.55 mmol) with Ba(OH)₂·8 H₂O (0.49 g, 1.55 mmol) in H₂O (5 ml) by extraction with Et₂O (5 × 10 ml). After concentration to 5–10 ml, the Et₂O soln. was stirred at 0° under SO₂ in a glass vessel. White crystals appeared immediately and were isolated by filtration: **12b** (0.306 g, 77%). M.p. 135° (subl.; EtOH/H₂O). IR (KBr): 3610, 3540, 2945, 2775, 2700, 2515, 1603, 1428, 1385, 1372, 1250, 1220, 1180, 1068, 1040, 850, 605. ¹H-NMR: Table 3. ¹³C-NMR (CD₃OD): 115.0 (Me₂C); 86.0 (C(3)); 82.6 (C(2)); 78.2 (C(1)); 62.9 (C(4)); 27.3, 25.1 (Me₂C); 16.2 (Me–C(4)). Anal. calc. for C₈H₁₅NO₃S·H₂O (255.27): C 37.49, H 6.68, N 5.46, S 12.51; found: C 37.7, H 6.7, N 5.7, S 12.6.

4-Amino-1,4-dideoxy-DL-erythrofurano-1-sulfonic Acid (13a). A soln. of **12a** (1.0 g, 4.48 mmol) in H₂O (10 ml) was stirred overnight at 50° under SO₂. At 0°, cold MeOH (20 ml) was added and the precipitate isolated: **13a** (0.52 g, 63%). Evaporation of the solvents gave a second crop of **13a** (0.14 g, 17%). White crystals. M.p. 170° (dec.; MeOH/H₂O). IR (KBr): 3510, 2980, 2770, 1710, 1590, 1428, 1300, 1205, 1175, 1128, 1000, 942, 813, 550. ¹H-NMR: Table 3. ¹³C-NMR (D₂O): 74.3 (C(2)); 73.7 (C(3)); 70.8 (C(1)); 51.2 (C(4)). Anal. calc. for C₄H₉NO₃S (183.13): C 26.23, H 4.95, N 7.65, S 17.51; found: C 26.1, H 5.1, N 7.6, S 17.5.

4-Amino-1,4,5-trideoxy-DL-ribofuranose-1-sulfonic Acid (13b). A soln. of **12b** (0.1 g, 0.39 mmol) in H₂O (3 ml) was stirred overnight at 35° under SO₂. After evaporation of H₂O, the residue was recrystallized in EtOH/H₂O to give anal. pure **13b** (30 mg, 40%). White crystals. M.p. > 160° (dec.; EtOH/H₂O). IR (KBr): 3460, 3000, 2960, 2780, 1600, 1240, 1222, 1185, 1045, 823, 610. ¹H-NMR: Table 3. ¹³C-NMR (D₂O): 75.9, 75.8 (C(2), C(3)); 72.4 (C(1)); 60.2 (C(4)); 15.3 (Me–C(4)). Anal. calc. for C₅H₁₁NO₃S (197.21): C 30.45, H 5.62, N 7.10, S 16.26; found: C 30.2, H 5.7, N 7.1, S 15.8.

8. *Cyano Derivatives. General Procedure*. To a HCN soln. in Et₂O (prepared from NaCN (0.3 g, 6 mmol, 6 equiv.) and a soln. of 2.1N HCl in Et₂O (2.2 ml, 4.5 equiv.) in Et₂O (1.5 ml) and H₂O (0.3 ml)) was added **11** (1 mmol) in Et₂O (1.5 ml). The soln. was stirred for 2 h at r.t. and then filtered and evaporated: pure **15**.

2,5-Imino-2,5-dideoxy-3,4-O-isopropylidene-DL-ribo-pentonitrile (15a). General Procedure with **11a** (94 mg, 0.67 mmol), HCl soln. in Et₂O (1.44 ml), and NaCN (0.2 g, 4 mmol) (0.11 g, 98%). Yellowish resin. IR (CHCl₃): 2254 (CN). ¹H-NMR: Table 3. ¹³C-NMR (CDCl₃): 117.9 (CN); 112.0 (Me₂C); 84.5 (C(3)); 82.7 (C(4)); 56.0 (C(2)); 52.5 (C(3)); 25.8, 24.0 (Me₂C). MS: 168 (6), 153 (31), 126 (78), 110 (40), 93 (54), 84 (35), 82 (40), 81 (40), 68 (13), 59 (34), 55 (63), 43 (100). HR-MS: 168.0895 (C₈H₁₂N₂O₂, calc. 168.0899).

2,5-Imino-2,5,6-trideoxy-3,4-O-isopropylidene-DL-allo-hexonitrile (15b). General Procedure with **11b** (0.148 g, 0.95 mmol), HCl soln. in Et₂O (2.04 ml), and NaCN (0.28 g, 5.7 mmol; 6 h at r.t.): **15b** (0.133 g, 77%). IR (CHCl₃): 2254 (CN). ¹H-NMR: Table 3. ¹³C-NMR (CDCl₃): 120.1 (CN); 112.8 (Me₂C); 86.7 (C(4)); 85.2 (C(3)); 61.0 (C(5)); 54.8 (C(2)); 26.5, 24.4 (Me₂C); 18.7 (Me–C(4)). MS: 182 (5), 167 (13), 124 (35), 107 (24), 101 (11), 95 (18), 91 (16), 82 (100), 69 (37), 55 (26). HR-MS: 182.1055 (C₉H₁₄N₂O₂⁺; calc. 182.1055).

9. *Amino Acids. General Procedure*. Nitrile **15** (1 mmol) was hydrolyzed in 6N HCl (1.5 ml) at 50°. The black soln. was neutralized with 2.5N NaOH and evaporated and the residue chromatographed (CHCl₃/MeOH/conc. NH₃ soln. 2:6:2).

(2RS,3SR,4RS)-3,4-Dihydroxyproline. General Procedure with **15a** (0.11 g, 0.65 mmol; overnight): **16a** (80 mg, 83%). Brown crystals. M.p. > 219° (H₂O/i-PrOH) ([12]: 241–242° (dec.)). IR (KBr): 3345, 3117, 2690, 2560, 2410, 1635, 1590, 1408, 1370, 1330, 1295, 1137, 1098, 1060; in good agreement with [12]. ¹H-NMR: 4.23 (m, H–C(3), H–C(4)); 3.86 (d, J = 5, H–C(2)); 3.42 (dd, J = 12, 5, H_a–C(5)); 3.18 (dd, J = 12, 4, H_b–C(5)). ¹³C-NMR (D₂O): 171.3, 74.0, 69.8, 64.2, 48.2; ¹H- and ¹³C-NMR in good agreement with [13a] [13b] for the (2R,3S,4R)- and (2S,3R,4S)-isomers.

(2RS,3SR,4RS,5RS)-3,4-Dihydroxy-4-methylproline (**16b**). General Procedure with **15b** (0.12 g, 0.65 mmol; 27 h). The collected crystals were recrystallized in H₂O/i-PrOH: pure **16b** (78 mg, 74%). Cream-colored crystals.

M.p. 115–116° (dec.). IR (KBr): 3307, 3060, 1620, 1569, 1440, 1405, 1384, 1325, 1281, 1140, 1070, 1040, 880, 665. ¹H-NMR (D₂O): 4.45 (dd, H–C(3)); 4.07 (d, H–C(2)); 3.94 (dd, H–C(4)); 3.69 (q, H–C(5)); 1.49 (d, Me–C(5)); J(2,3) = 2.4, J(3,4) = 4.1, J(4,5) = 8.4, J(5,Me) = 6.7. ¹³C-NMR (D₂O): 172.4 (CO₂H); 76.1 (C(4)); 74.3 (C(3)); 66.8 (C(2)); 58.0 (C(5)); 15.8 (Me–C(5)). HR-MS: 125.0473 (C₆H₇NO₂⁺, [M – 2 H₂O]⁺; calc. 125.0477).

10. *AIDS-Inhibition Assays*. The tests were made on aminosugars in C8166 cells infected with HIV-1 MN. Thus, 4 · 10⁴ cells were mixed with 5-fold dilutions of compounds prior to addition of virus (10, 50% cell culture infectious dose). Inhibition of infection was assessed by examining reduction of syncytia formation and by measuring virus yield after 5–6 days of incubation at 37°. Cell viability of infected and uninfected cells was determined by the XTT-formazan assay [27].

Virus yield was titrated on C8166 cells by doubling dilutions of the freshly collected supernatant from infected cells after 5–6 days of incubation.

REFERENCES

- [1] H. Paulsen, K. Propp, J. Brüning, *Chem. Ber.* **1969**, *102*, 469.
- [2] H. Paulsen, J. Brüning, K. Heyns, *Chem. Ber.* **1970**, *103*, 1621.
- [3] H. Paulsen, K. Steinert, K. Heyns, *Chem. Ber.* **1970**, *103*, 1599.
- [4] T. Shibata, O. Nakayama, Y. Tsurumi, M. Okuhara, H. Terano, M. Koksaka, *J. Antibiot.* **1988**, *41*, 296; H. Kayakiri, K. Nakamura, Sh. Takase, H. Setoi, I. Uchida, H. Terano, M. Hashimoto, T. Tada, Sh. Koda, *Chem. Pharm. Bull.* **1991**, *39*, 2807; H. Kayakiri, Sh. Takase, H. Setoi, I. Uchida, H. Terano, M. Hashimoto, *Tetrahedron Lett.* **1988**, *29*, 1725.
- [5] J. F. Witte, R. W. McClard, *Tetrahedron Lett.* **1991**, *32*, 3927.
- [6] B. A. Horenstein, R. F. Zabinski, V. L. Schramm, *Tetrahedron Lett.* **1993**, *34*, 7213.
- [7] Y. Arakawa, Sh. Yoshifuji, *Chem. Pharm. Bull.* **1991**, *39*, 2219.
- [8] A. Defoin, J. Pires, J. Streith, *Helv. Chim. Acta* **1991**, *74*, 1653.
- [9] Ch. A. Zezza, M. B. Smith, *J. Org. Chem.* **1988**, *53*, 1161.
- [10] a) G. W. Kirby, J. G. Sweeny, *J. Chem. Soc., Chem. Commun.* **1973**, 704; b) G. E. Keck, R. R. Webb, J. B. Yates, *Tetrahedron* **1981**, *37*, 4007; c) H.-Sh. Dang, A. G. Davies, *J. Chem. Soc., Perkin Trans. 2* **1991**, 721.
- [11] H. Böhshagen, W. Geiger, B. Junge, *Angew. Chem.* **1981**, *93*, 800.
- [12] C. B. Hudson, A. V. Robertson, W. R. J. Simpson, *Aust. J. Chem.* **1968**, *21*, 769.
- [13] a) P. D. Baird, J. C. Dho, G. W. J. Fleet, J. M. Peach, K. Prout, P. W. Smith, *J. Chem. Soc., Perkin Trans. 1* **1987**, 1785; b) G. W. J. Fleet, J. Ch. Son, *Tetrahedron* **1988**, *44*, 2637; c) N. Ikota, *Chem. Pharm. Bull.* **1993**, *41*, 1717.
- [14] C. J. Rule, B. A. Wurzburg, B. Ganem, *Tetrahedron Lett.* **1985**, *26*, 5379.
- [15] W. Kirmse, in 'Houben-Weyl, Methoden der organischen Chemie', 4th edn., G. T. Thieme Verlag, Stuttgart, 1972, Vol. 5/1b, p. 670.
- [16] V. Van Rheenen, R. C. Kelly, D. Y. Cha, *Tetrahedron Lett.* **1976**, *17*, 1973.
- [17] A. Defoin, H. Fritz, G. Geffroy, J. Streith, *Helv. Chim. Acta* **1988**, *71*, 1642.
- [18] H. Paulsen, F. Leupold, K. Todt, *Liebigs Ann. Chem.* **1966**, *692*, 200.
- [19] J. Firl, *Chem. Ber.* **1969**, *102*, 2169.
- [20] H. Paulsen, K. Todt, *Chem. Ber.* **1967**, *100*, 3385.
- [21] J. D. Stevens, H. G. Fletcher, Jr., *J. Org. Chem.* **1968**, *33*, 1799.
- [22] Y. Kodama, T. Tsuruoka, T. Niwa, Sh. Inouye, *J. Antibiot.* **1985**, *38*, 116.
- [23] A. Defoin, H. Sarazin, J. Streith, *Tetrahedron Lett.* **1993**, *34*, 4327.
- [24] B. D. Walker, M. Kowalski, W. Ch. Goh, K. Kozarsky, M. Krieger, C. Rosen, L. Rohrschneider, W. A. Haseltine, J. Sodroski, *Proc. Natl. Acad. Sci. U.S.A.* **1987**, *84*, 8120.
- [25] O. Bayer, in 'Houben-Weyl, Methoden der organischen Chemie', 4th edn., G. T. Thieme Verlag, Stuttgart, 1954, Vol. 7/1b, p. 118.
- [26] K. Akashi, R. E. Palermo, K. B. Sharpless, *J. Org. Chem.* **1978**, *43*, 2063.
- [27] O. S. Weislow, R. Kiser, D. L. Fine, J. Bader, R. H. Shoemaker, M. R. Boyd, *J. Natl. Cancer Inst.* **1989**, *81*, 577.